

A FLAVONOL GLYCOSIDE FROM *EPIMEDIUM DIPHYLLUM*

MIZUO MIZUNO,* MUNEKAZU IINUMA, TOSHIYUKI TANAKA, NORIO SAKAKIBARA, MASATOSHI NISHI,†AKIRA INADA† and TSUTOMU NAKANISHI†

Department of Pharmacognosy, Gifu Pharmaceutical University, 6-1 Mitahora-higashi 5 chome, Gifu 502, Japan; †Faculty of Pharmaceutical Sciences, Setsunan University, 45-1 Nagaotoge-cho, Hirakata, Osaka 573-01, Japan

(Received 3 January 1989)

Key Word Index—*Epimedium diphyllosum*; Berberidaceae; flavonol glycoside; diphyllside C.

Abstract—A novel flavonol glycoside named diphyllside C was isolated from the underground parts of *Epimedium diphyllosum* and its structure was determined on the basis of spectral analyses (negative ion FAB-MS, ^1H - ^1H COSY, NOESY, INEPT and ^1H - ^{13}C COSY, etc) as des-*O*-methylanhydroicaritin 3-*O*- β -D-glucosyl-(1 \rightarrow 2)- α -L-rhamnoside 7-*O*- β -D-glucosyl-(1 \rightarrow 2)- β -D-glucoside.

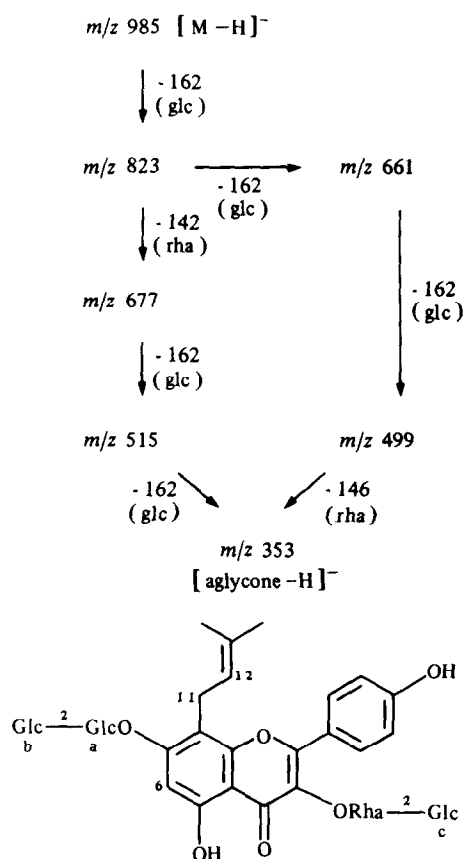
INTRODUCTION

In our previous papers the isolation and structural elucidation of four new flavonol glycosides were reported from *Epimedium sagittatum*: icaritin 3-*O*- α -rhamnoside [1] and sagittatosides A–C [2] and two: diphyllsides A and B from *E. diphyllosum* [3]. In the course of our search for bioactive principles in *Epimedium* species as well as a chemotaxonomic investigation of the genus *Epimedium*, further constituents of the underground parts of *E. diphyllosum* (Morr. et Decne) Lodd. (Berberidaceae) were investigated. The present paper deals with the structure of a novel flavonol glycoside with different disaccharides at C-3 and C-7.

RESULTS AND DISCUSSION

After repeated silica gel chromatography of the butanol-soluble portion of a 70% methanolic extract, and final purification by medium pressure liquid chromatography, diphyllside C (**1**), mp 184–185°, $[\alpha]_D^{25}$ –76.3° (MeOH; *c* 0.25), was obtained as a pale yellow powder. The UV spectrum showed absorption bands at 270, 320 and 353 nm. Bathochromic-shifts on addition of some reagents indicated the presence of free hydroxyl groups at C-5 and C-4' of the flavonol skeleton. In the ^1H NMR spectrum (all protons were assigned by ^1H - ^1H COSY, NOESY and ^1H - ^{13}C COSY) one-proton multiplets at δ 3.61 and 3.47, a one-proton multiplet at 5.14 in addition to two three-proton singlets at 1.62 and 1.69 showed the presence of a γ,γ -dimethylallyl group. This was also supported by signals at 21.25 (triplet) assigned to C-11, 122.56 (doublet) to C-12, 130.59 (singlet) to C-13, and 25.41 and 17.81 (both doublets) to C-14 and C-15 in the ^{13}C NMR spectrum, in which all carbons were assigned by INEPT and ^1H - ^{13}C COSY. The B ring moiety was oxygenated only at C-4' on account of the signals of A_2B_2 at δ 7.81 and 6.95. On the basis of significant fragment

ion at m/z 353 in the FAB-MS, the aglycone moiety of **1** was identified as des-*O*-methylanhydroicaritin (= 8- γ,γ -dimethylallyl-3,5,7,4'-tetrahydroxyflavone). Negative FAB-MS gave fragment ions which indicated that



1 Diphyllside C

* Author to whom correspondence should be addressed.

1 possessed three hexoses and a methyl pentose.

Acetylation of **1** gave the corresponding pentadeca-acetate (**2**), $C_{74}H_{88}O_{40}$, as a colourless powder. In the negative FAB-MS of **2** a $[M-H]^-$ ion was observed at m/z 1615. Further proof of the complete structure of **1** was obtained from the 1H NMR of **2** and with the aid of 1H - 1H COSY, NOESY and 1H - ^{13}C COSY all the protons of **2** have been reasonably assigned, the data for which are shown in Table 1. A consideration of the chemical shifts and the coupling constants of four anomeric protons [δ 4.60 ($J = 8.0$ Hz), 4.77 ($J = 8.0$ Hz), 5.08 ($J = 7.5$ Hz) and 5.61 ($J = 1.8$ Hz), each doublet] and all the other sugar proton signals suggested the presence of two terminal β -D-glucopyranoses (C1 conformation) (hereafter shown as glc-b and glc-c) and two inner sugars, i.e. α -L-rhamnopyranose (1C conformation) and β -D-glucopyranose (C1 conformation) (shown as glc-a). Furthermore, it was also inferred that each of the inner sugars is linked at the 2β -OH groups with a terminal glucosyl moiety. The final structure for **1** was established by NOESY experiments of **1** and **2** as follows: (i) cross peaks were observed between the anomeric proton of rhamnose and the protons at C-2' and C-6' of the aglycone, and also between the 2β -proton of rhamnose and the anomeric proton of a terminal glucose (glc-c), indicating that a β -D-glucosyl-(1 \rightarrow 2)- α -L-rhamnosyl residue is connected with a hydroxyl group at C-3 of the aglycone; (ii) NOE effects were observed between the anomeric proton of inner glucose (glc-a) and the proton at C-6 and also between the 2β -proton of inner glucose and the anomeric proton of the other terminal glucose (glc-b). This finding showed that a β -D-glucopyranosyl-(1 \rightarrow 2)- β -D-glucopyranosyl residue is linked with a hydroxyl group at C-7. These two disaccharide structures at C-3 and C-7 were further corroborated by the following alternative investigation; (i) in the 1H NMR experiments, both the 2β -protons of rhamnose and the inner glucose (glc-a) in **2** resonated at almost the same field as those in the original glycoside; (ii) in the ^{13}C NMR spectrum of **1**, the carbons at C-2 of rhamnose resonated at δ 81.21, which is a shift upfield by ca 10 ppm more than that of a usual rhamnose. Analogously, the C-2 signal of an inner glucose (glc-a) appeared at δ 80.94, a shift of 6–7 ppm upfield in comparison with the terminal glucoses glc-b and glc-c; (iii) the fissions in the negative ion FAB-MS of **1** were reasonably consistent with this bisdemioside structure. From the above spectral evidence **1** is concluded to be des-*O*-methylanhydrocaritin 3-*O*- β -D-glucosyl-(1 \rightarrow 2)- α -L-rhamnoside 7-*O*-

β -D-glucopyranosyl-(1 \rightarrow 2)- β -D-glucoside. Compound **1** can also be regarded as diphyllloside A [des-*O*-methylanhydrocaritin 3-glucosyl-(1 \rightarrow 2)-rhamnoside 7-glucoside] with an extra glucose at C-7. It should be pointed out that diphyllloside A was also reported by Fukai and Nomura [4] as a new compound from *E. grandiflorum* at the same time as our previous publication [3] and they named it ikarisoside C.

EXPERIMENTAL

Plant material and procedures of extraction and isolation were described in ref. [3].

Compound 1 (diphyllloside C). A yellow powder (EtOH-H₂O) (35 mg) was obtained, mp 184–185°, $C_{44}H_{58}O_{25}$, $[\alpha]_D -76.3^\circ$ (MeOH; c 0.25). IR ν_{max}^{KBr} cm^{-1} : 3380, 2920, 1650, 1600. UV λ_{max}^{MeOH} nm: 221sh, 270, 320, 353, + NaOMe: 250, 272, 386, + AlCl₃: 241, 275sh, 305, 360, + AlCl₃-HCl: 241, 275sh, 305, 360, + NaOAc: 271, 322, 360, + NaOAc-H₃BO₃: 271, 322, 350sh. EIMS (m/z) (rel. int.): 354 (22), 339 (21), 299 (16), 165 (4), 121 (13). 1H NMR (400 MHz, DMSO- d_6) δ : [aglycone moiety] 7.81 (d , $J = 8.5$ Hz, H-2', 6'), 6.95 (d , $J = 8.5$ Hz, H-3', 5'), 6.62 (s , H-6), 3.6] and 3.47 (m , H₂-11), 5.14 (m , H-12), 1.62 and 1.69 (each s , Me), [sugar moieties] (glc-a) 5.18 (d , $J = 7.8$ Hz, H-1), 3.67 (dd , $J = 7.8$ Hz, H-2); (glc-b) 4.59 (d , $J = 7.8$ Hz, H-1), 3.03 (dd , $J = 7.8$, 7.8 Hz, H-2); (rha) 5.59 (s , H-1), 4.13 (d , $J = 3.3$ Hz, H-2), 0.88 (d , $J = 5.7$ Hz, H-6); (glc-c) 4.27 (d , $J = 7.8$ Hz, H-1), 3.00 (dd , $J = 7.8$, 7.8 Hz, H-2). ^{13}C NMR (100.5 MHz, DMSO- d_6) δ : [aglycone moiety] 152.87 (s , C-2), 134.49 (s , C-3), 178.11 (s , C-4), 159.88 (s , C-5), 97.65 (d , C-6), 160.23 (s , C-7), 108.23 (s , C-8), 157.33 (s , C-9), 105.37 (s , C-10), 21.25 (t , C-11), 122.56 (d , C-12), 130.59 (s , C-13), 25.41 and 17.81 (each d , C-14 and C-15), 120.29 (s , C-1'), 130.52 (d , C-2', 6'), 115.36 (d , C-3', 5'), 158.91 (s , C-4'), [sugar moieties] (glc-a) 98.19 (d , C-1), 80.94 (d , C-2); (glc-b) 103.93 (d , C-1), 74.61 (d , C-2); (rha) 100.85 (d , C-1), 81.21 (d , C-2), 70.34 (d , C-3), 71.55 (d , C-4), 70.05 (d , C-5), 17.26 (q , C-6); (glc-c) 106.06 (d , C-1), 73.73 (d , C-2); other glucosyl carbons 76.53, 76.13, 76.13 ($3 \times$ C-3), 69.43, 69.19, 69.08 ($3 \times$ C-4), 76.93, 76.89, 76.89 ($3 \times$ C-5), 60.47, 60.34, 60.25 ($3 \times$ C-6).

Compound 2 (pentadeca-acetate of **1**): A colourless powder, $C_{74}H_{88}O_{40}$, $[\alpha]_D -71.1^\circ$ (CHCl₃; c 0.33). IR ν_{max}^{KBr} cm^{-1} : 1750 (Ac), 1630 (C=O), 1605 (C=C). 1H NMR (400 MHz, CDCl₃) δ : [aglycone moiety] 7.86 (d , $J = 8.7$ Hz, H-2', 6'), 7.26 (d , $J = 8.7$ Hz, H-3', 5'), 6.80 (s , H-6), 5.17 (dd , $J = 5.0$, 7.5 Hz, H-12), 3.68 (d , $J = 5.0$, 7.5 Hz, H-11), 3.59 (d , $J = 7.5$, 14.0 Hz, H-11), 1.67 and 1.73 (each s , Me) [acetyl methyl groups] 2.38, 2.33, 2.18, 2.14, 2.10, 2.07 ($\times 2$), 2.06, 2.05, 2.03, 2.02, 2.01 ($\times 2$), 1.99, 1.97 [sugar moiety] shown in Table 1.

Table 1. 1H NMR spectral data of the sugar moiety of the pentadeca-acetate of **1** (**2**)

	glc-a	glc-b	glc-c	rha
1	5.08 (d , $J = 7.5$ Hz)	4.77 (d , $J = 8.0$ Hz)	4.60 (d , $J = 8.0$ Hz)	5.61 (d , $J = 1.8$ Hz)
2	4.12 (dd , $J = 7.5$, 9.3 Hz)	4.93 (dd , $J = 8.0$, 9.3 Hz)	5.03 (dd , $J = 8.0$, 9.3 Hz)	4.43 (dd , $J = 1.8$, 3.5 Hz)
3	5.29 (dd , $J = 9.3$, 9.3 Hz)	5.15 (dd , $J = 9.3$, 9.3 Hz)	5.22 (dd , $J = 9.3$, 9.3 Hz)	5.20 (dd , $J = 3.5$, 10.0 Hz)
4	5.06 (dd , $J = 9.3$, 9.3 Hz)	4.94 (dd , $J = 9.3$, 9.3 Hz)	5.10 (dd , $J = 9.3$, 9.3 Hz)	4.85 (dd , $J = 10.0$, 10.0 Hz)
5	3.92 (ddd , $J = 2.5$, 6.0, 9.3 Hz)	3.68 (ddd , $J = 2.5$, 3.5, 9.3 Hz)	3.71 (ddd , $J = 2.5$, 3.8, 9.3 Hz)	3.24 (dq , $J = 10.0$, 6.0 Hz)
6	4.26 (dd , $J = 6.0$, 12.0 Hz)	4.18 (dd , $J = 3.5$, 12.0 Hz)	4.36 (dd , $J = 3.8$, 12.0 Hz)	0.84 (d , $J = 6.0$ Hz)
	4.17 (dd , $J = 2.5$, 12.0 Hz)	4.08 (dd , $J = 2.5$, 12.0 Hz)	4.06 (dd , $J = 2.5$, 12.0 Hz)	

REFERENCES

1. Mizuno, M., Hanioka, S., Suzuki, N., Iinuma, M., Tanaka, T., Liu, X. and Min, Z. (1987) *Phytochemistry* **26**, 861.
2. Mizuno, M., Sakakibara, N., Hanioka, S., Iinuma, M., Tanaka, T., Liu, X. and Shi, B. (1988) *Phytochemistry* **27**, 3641.
3. Mizuno, M., Iinuma, M., Tanaka, T., Sakakibara, N., Fujikawa, T., Hanioka, S., Ishida, Y., Liu, X. and Murata, H. (1988) *Phytochemistry* **27**, 3645.
4. Fukai, T. and Nomura, T. (1988) *Phytochemistry* **27**, 259.

Phytochemistry, Vol. 28, No. 9, pp. 2529–2530, 1989.
Printed in Great Britain.

0031-9422/89 \$3.00 + 0.00
© 1989 Maxwell Pergamon Macmillan plc.

TWO ANTHOCHLOR PIGMENTS FROM HEARTWOOD OF *PTEROCARPUS MARSUPIUM*

POONAM MOHAN and T. JOSHI

Department of Chemistry, University of Allahabad, 211 002, India

(Received 26 January 1989)

Key Word Index—*Pterocarpus marsupium*; Leguminosae; 6,4'-dihydroxy-7-methylaurone 6-*O*-rhamnopyranoside; aureusidin 6-*O*-rhamnopyranoside.

Abstract—Two new aurone glycosides, 6,4'-dihydroxy-7-methylaurone 6-*O*-rhamnopyranoside and 4,6,3',4'-tetrahydroxy aurone 6-*O*-rhamnopyranoside have been isolated and identified from the heartwood of *Pterocarpus marsupium*.

From the aqueous extract of the heartwood of *P. marsupium*, two novel aurone glycosides have been isolated and identified. Compound **1**, the yellow pigment analysed for $C_{22}H_{22}O_8$, mp 170°. It was found to be glycosidic in nature [1]. On acid hydrolysis (7% H_2SO_4) it gave an aglycone and rhamnose, identified by co-chromatography with an authentic sample and by 1H NMR spectral analysis of the glycoside (a doublet at δ 1.20 corresponding to the three protons of rhamnosyl—Me group, broad signal at δ 3.5–3.82 for four sugar protons and a singlet at δ 4.2 due to C-1" proton of rhamnose).

The aglycone, $C_{16}H_{12}O_4$ was characterized as an aurone on the basis of colour reactions [2] and UV spectral data [3]. 1H NMR studies showed six aromatic protons suggesting a trisubstituted aurone. A multiplet at δ 7.69–7.9 (2H) was due to C-2' and -6' and multiplet at δ 6.8–7.0 (2H) for C-3' and -5' protons. There was a singlet at δ 6.67 for benzylic proton (=CH–) [4]. A singlet at δ 1.44 was assignable to 3H of the –Me group. On acetylation it gave a diacetate, mp 96°, showing the presence of two hydroxyl groups. The positions of hydroxyls were shown to be at C-4' and C-6 of aglycone by UV spectral shifts (bathochromic shift of 45 and 46 nm of band 1 with sodium methoxide and sodium acetate respectively). The presence of a free –OH at C-4' position in the glycoside was confirmed by a large bathochromic shift in λ_{max}^{MeOH} (ca 70 nm) upon addition of sodium methoxide and (68 nm) with sodium acetate. C-7 position of –Me group was confirmed by NMR.

Mass spectral data showed a molecular ion peak at 414. Two fragments at m/z 150 and m/z 118 showed that

one hydroxyl group was present in the B ring and –Me was present in the A ring. Thus the structure of **1** is confirmed. This compound has not been reported earlier from any other plant source.

Compound **2**, a yellow crystalline compound analysed for $C_{21}H_{20}O_{10}$, mp 264°, was found to be glycosidic in nature [1]. On hydrolysis it gave rhamnose (co-PC) and an aglycone which was shown to be an aurone by its colour reactions [2] and UV spectrum [3]. 1H NMR studies of the aglycone showed five aromatic protons suggesting a tetrasubstituted nucleus. On acetylation it gave a tetraacetate, mp 182°, showing the presence of four hydroxyl groups. The aglycone was found to be the same as a [synthetic] sample of aureusidin (4,6,3',4'-tetrahydroxyaurone) on the basis of chromatography and spectroscopy [5, 6]. The UV spectrum of the glycoside showed that the sugar was not at C-4 (a large bathochromic shift of 60 nm by aluminium chloride) or C-4' (bathochromic shift of 85 nm in alkali). Methylation of the glycoside, followed by acid hydrolysis gave a product which was chromatographically and spectrally identical with a [synthetic] sample of 6-hydroxy-4,3',4'-trimethoxyaurone. Thus the structure of **2** is confirmed; this is a new glycoside but the aglycone has been reported earlier [5, 6].

The structures of aglycones were further confirmed by synthesis. Aglycone of **1** was synthesized by condensing 6-hydroxy-7-methylcoumaranone and *p*-hydroxybenzaldehyde. Aureusidin, the aglycone of **2** was prepared from 4,6-dihydroxycoumaranone and 3,4-dihydroxybenzaldehyde.